

# M23 isoform is superior to M1 for detecting AQP4-IgG seroreversion during NMOSD

Vinícius de Oliveira Boldrini<sup>1\*</sup>, Leonie Erpenbeck<sup>1\*</sup>, Elif Dönmez<sup>1</sup>, Tania Kümpfel<sup>1#</sup>, Simone Mader<sup>1#</sup>, Edgar Meinl<sup>1#</sup>

<sup>1</sup> Institute of Clinical Neuroimmunology, Biomedical Center (BMC), LMU Munich, 82152 Martinsried, Germany

Correspondence: T.K. (Tania.Kuempfel@med.uni-muenchen.de), S.M. (Simone.Mader@med.uni-muenchen.de), E.M. (Edgar.Meinl@med.uni-muenchen.de)

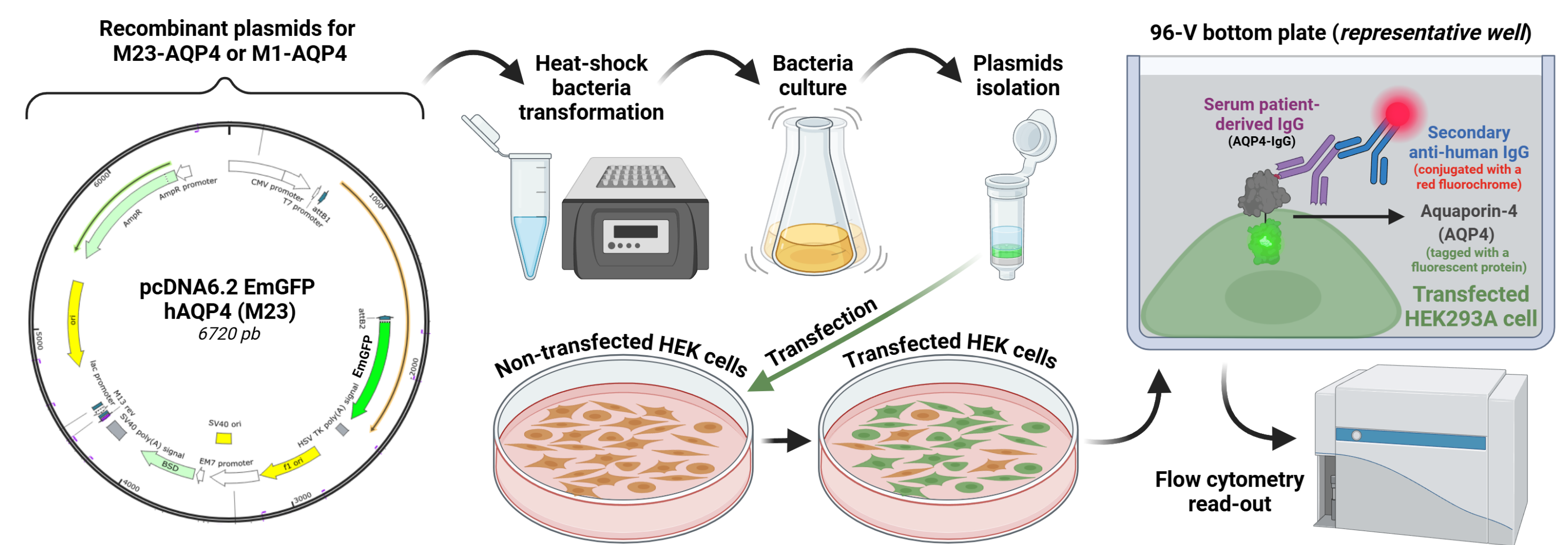
## Abstract

Antibodies against AQP4 (AQP4-IgG) are present in more than 80% of patients with Neuromyelitis Optica spectrum disorder (NMOSD) and play an important role in disease pathophysiology. Lesions in AQP4-positive NMOSD patients are characterized by mixed immune cell infiltrates as well as IgG and complement deposition around the blood vessels<sup>1,2,3</sup>.

Importantly, and described in detail in recent papers, some patients with AQP4-positive NMOSD may experience fluctuations in serum AQP4-IgG levels, including seroreversion (loss of detectable antibodies) and eventual reconversion (the reappearance of detectable antibodies) during the disease course. The clinical relevance of these processes is still under debate<sup>4,5</sup>.

Here, we investigated the potential differences in defining seropositivity, seroreversion and reconversion based on the detection of AQP4-IgG to the M23- and M1- AQP4 isoforms in NMOSD patients.

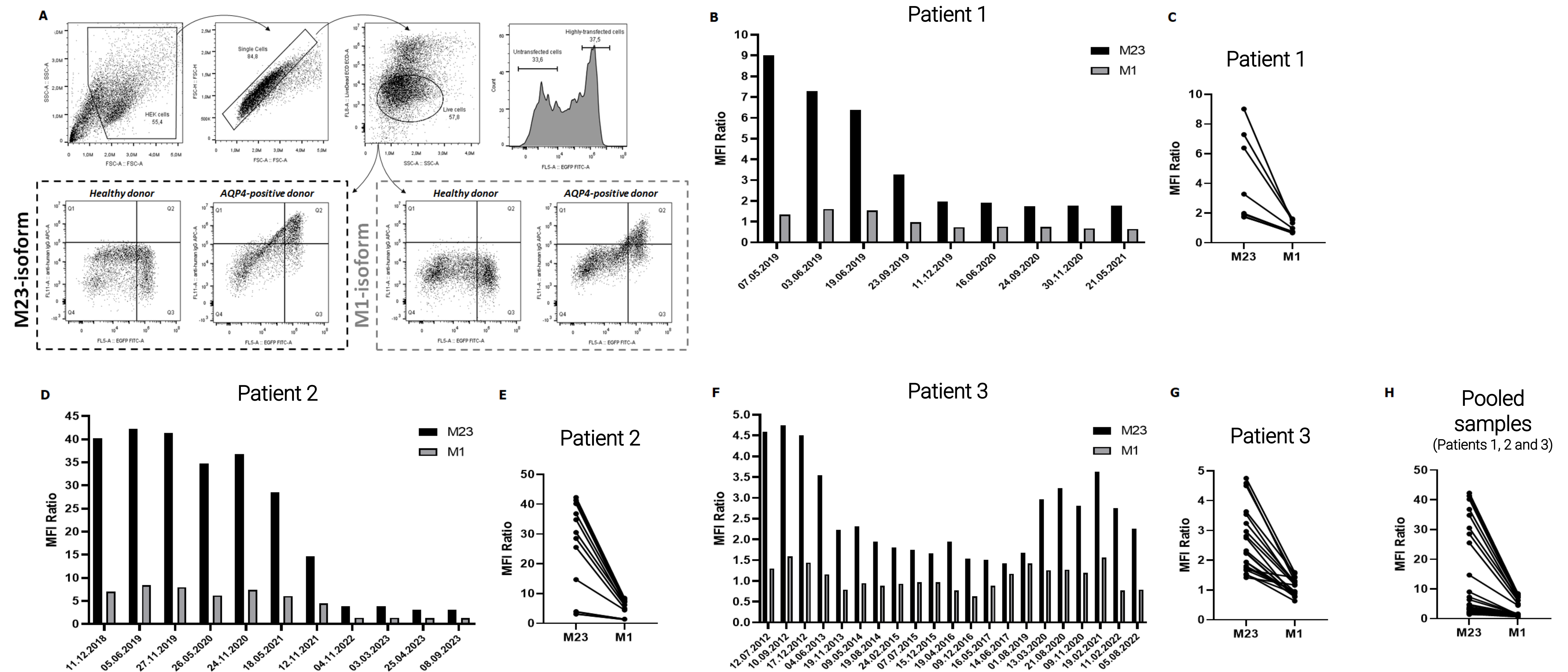
## Methods



**Figure 1.** Methods for obtaining M23-AQP4 and M1-AQP4 transfected cells for the investigation of AQP4-IgG in serum samples using the live cell-based assay (CBA).

Human embryonic kidney cells (HEK293A) were used for the transient transfection with the plasmids pcDNA6.2 EmGFP hAQP4 (M23) and pcDNA6.2 EmGFP hAQP4 (M1). Serum samples collected longitudinally from 3 AQP4-positive NMOSD donors (below) were diluted 1:50 and incubated with the transfected cells, followed by anti-human IgG secondary antibody for detecting cell-bound AQP4-IgG. Samples were acquired through flow cytometry (FACS-CBA).

## Results



**Figure 2.** Longitudinal assessment of M23-AQP4-IgG and M1-AQP4-IgG by FACS-CBA

(A) Gate strategy for flow cytometry analysis determined two populations of cells gated based on their intensity of emerald green fluorescent protein (EmGFP) expression: positive (highly-transfected for AQP4) and negative (untransfected for AQP4). The mean Alexa Fluor 647 fluorescence intensity (MFI) for the EmGFP-positive population/EmGFP-negative population was calculated as a surrogate parameter for AQP4-specific IgG reactivity to the M23 and M1 isoforms, respectively. (B and C) Patient 1 – Sex: female; Date of birth: 1982; First clinical manifestation: 2005; First positive test record: 2018; Treatment: Rituximab; Samples collected over 745 days, Interquartile range (IQR) = 505 days [Q3 (540) – Q1 (35)]. (D and E) Patient 2 – Sex: male; Date of birth: 1959; First clinical manifestation: 1999; First positive test record: 2012; Treatment sequence: Cyclophosphamide, Azathioprine, Cyclophosphamide, Plasmapheresis, Rituximab, Satralizumab; Samples collected over 1732 days, IQR = 1192 [1543 – 351]. (F and G) Patient 3 – Sex: female; Date of birth: 1964; First clinical manifestation: 1996; First positive test record: 2012; Treatment: Azathioprine, Cyclosporine A, Azathioprine, Cyclophosphamide, Mycophenolate mofetil; Samples collected over 3388 days, IQR = 2301 [2881.5 – 580.5]. (H) Pooled MFIs for the three donors.

## Conclusion

Larger cohorts may provide more powerful insights into whether testing for the M23-AQP4 isoform using FACS-CBA demonstrates higher sensitivity than M1-AQP4 in measuring longitudinal fluctuations of serum AQP4-IgG. This should be considered for accurately examining the phenomenon of seroreversion/reconversion observed in some AQP4-positive NMOSD patients.

## References

- Krumbholz, M. & Meinl, E. B cells in MS and NMO: Pathogenesis and therapy. *Semin. Immunopathol.* **36**, 339–350 (2014).
- Mader, S., Kümpfel, T. & Meinl, E. Pathomechanisms in demyelination and astrocytopathy: autoantibodies to AQP4, MOG, GFAP, GRP78 and beyond. *Curr. Opin. Neurol.* **35**, 427–435 (2022).
- Kümpfel, T. *et al.* Update on the diagnosis and treatment of neuromyelitis optica spectrum disorders (NMOSD) – revised recommendations of the Neuromyelitis Optica Study Group (NEMOS). Part II: Attack therapy and long term management. *J. Neurol.* **27**, 141–176 (2024).
- Majed, M. *et al.* Alterations in Aquaporin-4-IgG Serostatus in 986 Patients: A Laboratory-Based Longitudinal Analysis. *Ann. Neurol.* **94**, 727–735 (2023).
- Yin, H. X. *et al.* AQP4 Antibody Dynamics and Relapse Risk in Seropositive Neuromyelitis Optica Spectrum Disorder Treated with Immunosuppressants. *Ann. Neurol.* **93**, 1069–1081 (2023).

## Funding and Acknowledgement

V.O.B. was nominated Young Investigator (2021), awarded the Fellow Community Small Grant Fund (2022), and appointed as a member of the Steering Committee for Young Investigators (2024) from the European Charcot Foundation (ECF).

V.O.B. was named The Sumaira Foundation (TSF) Ambassador for Brazil and Germany (2024). V.O.B. was awarded the CAPES-Alexander von Humboldt (AvH) postdoctoral fellowship (2023) – call 19/2023 – to be held under the supervision of Prof. Edgar Meinl, MD (Human Neuroimmunology Lab.), and P.D. Simone Mader (CNS-reactive antibodies Lab.) at the Institute of Clinical Neuroimmunology, Ludwig-Maximilians-Universität - München (LMU München).