

## Cytotoxic B cells in MS patients

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## BACKGROUND

During relapsing-remitting MS (RRMS), CD8+ T lymphocytes infiltrate into the central nervous system (CNS) being found close to oligodendrocytes and neurons. Moreover, in severe/fatal MS relapses a massive infiltration of CD8<sup>+</sup> T cells expressing granzyme B (GzmB) was described in CNS parenchyma, evidencing an aberrant cytotoxic behavior. At the same time, B cells and other subsets were demonstrated to share cytotoxic behavior in several diseases. Since cytotoxicity is thought to be a central mechanism for neurodegeneration, we intend to investigate whether "cytotoxic" B cells occur in RRMS patients.

## **METHODS**

**104 RRMS patients** (19 Untreated, 15 Glatiramer Acetate [GA], 24 Interferon- $\beta$  [IFN], 14 Fingolimod [FTY] and 32 Natalizumab [NTZ]), according to the McDonald criteria were recruited in Neurology Clinic at University of Campinas Hospital (UNICAMP). Also, **58 healthy subjects** were included in the control groups. All subjects signed a term of consent approved by the University Committee for Ethical Research (CAAE: 53022516.3.0000.5404).



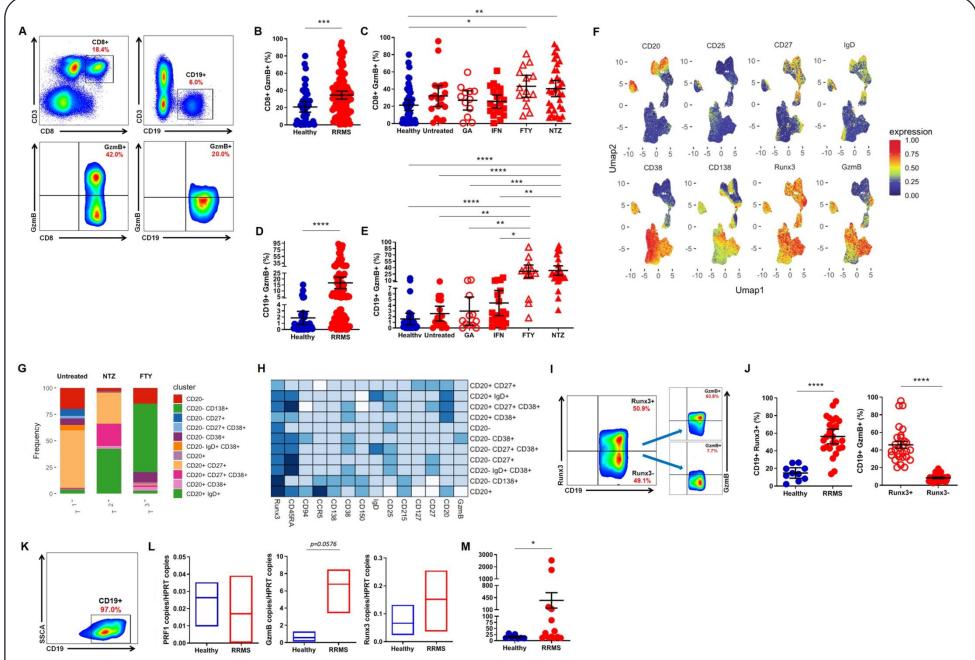


Figure 1: GzmB-expressing B cells in MS patients. (A) Gate strategy for CD8<sup>+</sup>GzmB<sup>+</sup>T lymphocytes and CD19<sup>+</sup>GzmB<sup>+</sup> B cells. (B) Proportion (%) of CD8<sup>+</sup>GzmB<sup>+</sup>T lymphocytes in reated RRMS patients (*GA*, *IFN*, *FTY*, *NTZ*) (red), untreated MS (red) and healthy donors (blue). (D) Proportion (%) of CD19<sup>+</sup>GzmB<sup>+</sup> B cells in RRMS patients (*GA*, *IFN*, *FTY*, *NTZ*) (red), untreated MS (red) and healthy donors (blue). (D) Proportion (%) of CD19<sup>+</sup>GzmB<sup>+</sup> B cells in RRMS patients (*GA*, *IFN*, *FTY*, *NTZ*) (red), untreated MRS (red) and healthy donors (blue). (E) Proportion (%) of CD19<sup>+</sup>GzmB<sup>+</sup> B cells in treated RRMS patients (*GA*, *IFN*, *FTY*, *NTZ*) (red), untreated RRMS (red) and healthy donors (blue). (F) UMAP gated in CD19<sup>+</sup> B cells from RRMS patients with different conditions non identified and based on the arcsine-transformed expression of the markers. (G) Barplot representing the frequency of each subpopulation in CD19<sup>+</sup> B cells. (H) Heatmap of the expression of the markers in subpopulations manually identified in CD19<sup>+</sup> B cells. (I) Gate strategy for GzmB-derived CD19<sup>+</sup>Runx3<sup>+</sup> B cells. (J) Proportion (%) of CD19<sup>+</sup>Runx3<sup>+</sup> B cells in RRMS patients (red) and healthy donors (blue) and proportion (%) of GzmB-derived CD19<sup>+</sup>Runx3<sup>+</sup> B cells in RRMS patients (red) and healthy donors (blue). (M) Concentration (pg/mL) of GzmB-derived from CD19<sup>+</sup> B cells supernatants from RRMS patients (red) and healthy donors (blue). (M) Concentration (pg/mL) of GzmB-derived from CD19<sup>+</sup> B cells supernatants from RRMS patients (red) and healthy donors (blue). \**p<0.05*, \*\**p<0.01*, \*\*\**p<0.001*, \*\*\**p<* 

## CONCLUSION

Increased percentage of CD19<sup>+</sup>GzmB<sup>+</sup> B cells was observed in FTY and NTZ subgroups when compared to GA, IFN and untreated RRMS patients. Moreover, using high-dimensional FACS (UMAP), we observed that CD19<sup>+</sup>CD138<sup>+</sup> plasma cells, but not CD19<sup>+</sup>CD20<sup>+</sup> B cells, seem to represent a main subset of B cells involved in GzmB-expression. Also, Runx3, a master regulator of cytotoxic activity classically described in CD8<sup>+</sup> T lymphocytes, seems to be associated with GzmB expression in CD19<sup>+</sup> B cells in MS patients. Thus, in addition to antigen presentation and cytokine production, ectopic cytotoxicity may represent a novel antibody-independent mechanism derived from B cells with possible implications for MS pathophysiology. Further investigations, in larger cohorts, may elucidate the eventual clinical/therapeutic relevance of "cytotoxic" B cells during MS.